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SCREENING OF 166 ANTIFERTILITY MEDICINAL PLANTS: REVIEW

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ABSTRACT

Medicinal plants are the valuable sources of herbal contraceptive well known to the Ancient Physicians of India. Modern contraceptives are restricted among women due to its unwanted side effects. The medical scientists are in search for suitable and active antifertility agents of synthetic and herbal origin to prevent conception. This review reveals with the number of herbal plants with their chemical constituents. Overall, 200 articles were reviewed and out of this 166 articles of proved effect of plants or isolated constituents in laboratory animals against antifertility were referred for citation. We found that all the reported extracts and individual compounds from these natural plants are beneficial in antifertility. This review indicates that it is time to increase the experimental studies to discover new chemical entities from vast unexploited plants having potential role in antifertility and also the exploration of mechanisms of action.

INTRODUCTION

Rapidly growing population is the most explosive problem being faced all around the world in developing countries like India and China where poverty is touching new heights. According to United nation projects that the population hit 6.5 billion by 2020 and 8.2 billion by 2050 in developed country. This steady rise in it causes detrimental impact on the international economy [1]. To combat this situation there is a need to look for a way of controlling the population growth. Fertility regulation has become the major concern of people of all walks of life [2]. Several remarkable steps have been taken by the Government of India and international bodies including NGOs in this direction to prevent conception viz. Popularizing family planning programmed public awareness and free distribution of contraceptives etc [3, 4]. India was the first country that launched “National Family Planning Program” in 1952 for the stabilization of population. The objective of this programme was “reduced birth rate to the extent essential to stabilize the population at a level consistent with the requirement of the national economy [5]. In olden times traditional methods are adopted that provided numerous ideas for reliable contraceptives of today’s including the birth controlling pills, Copper bearing Intrauterine devices, Condoms, Diaphragm were intimately related to sexual intercourse therefore disliked by most couples and thus they have higher failure rates [6]. Nowadays hormonal contraceptives are widely used with 100% confidence and results in an unacceptable rate of unwanted pregnancies and other side effects [7]. The common side effects are breast cancer, cervical cancer, gastric trouble, obesity, venous thromboembolism etc. Keeping these things in mind there is a search for a suitable product from traditionally used indigenous medicinal plants and a number of plants had been screened in attempt to replace steroidal contraceptives and found to be safe and effective antifertility agents [8, 9].

Herbals as antifertility agents

Herbal medicines had become an integral part of life in many communities and considered as a promising avenue for the discovery of new drugs due to its easy access and relatively low cost. Medicines derived from herbal drugs are used as indigenous cure in folklore or traditional systems of medicine. As per estimation of World Health Organization that 80% of people worldwide rely on herbal medicines for some part of their primary health care [10]. India has often been referred to as the medicinal garden of the world. About 45,000 plant species have been claimed to possess medicinal properties. Ancient knowledge coupled with scientific principles can come to the forefront and provide us with powerful remedies to eradicate the

diseases [11]. A brief description of plants having antifertility activity with their active constituents is given in table 1. From the study of all antifertility medicinal plants, it was concluded that the potential of activity order of different parts is Leaf>Seed>whole plant>root>Aerial part=Bark>Stem>Fruit=Flower>Tuber>Stem bark>Rhizome. Leaves have maximum and rhizome have minimum potential for antifertility activity (figure 2).

Common antifertility animal models used for listed medicinal plants

Antifertility activity in herbals is evaluated with different animal models using various parameters like antagonism of uterus weight, antagonism of the effect of testosterone on weight of ventral prostate, seminal vesicles and musculus levator ani., anti androgenic activity in female rats, anti androgenic activity on sebaceous glands, test for anti- androgenic activity, test for anti-estrogen, test for an ovulatory agents, ovum count etc [184, 185].

CONCLUSION

In the present review, we have made an attempt to provide detail information and idea about the significance of herbal drugs towards antifertility activity and this may focus researcher's attention for clinical studies which would be of great scientific contribution to the society.

ACKNOWLEDGEMENT

No conflict of interest

Table: 1List of plants having antifertility activity with their active constituents

Botanical name	Family	Parts used	Active constituents	Uses	Reference
<i>Abrus precatorius</i> L.	Leguminosae	Seed	PEP-103, PEP-104, Abridine	Used as oral contraceptive, prevents implantation of fertilized ovum by inhibiting endometrial alkaline phosphate. Sperm anti-motility activity.	[12,13,14]
<i>Acacia arabica</i>	Fabaceae	Stem bark	-	Effective oral contraceptive in rats and inhibits implantation	[15]
<i>Acacia caesia</i> (wight & Arn.)	Mimosaceae	Bark	Acacia acid saponin, Lupeol, α -spinosterol	Spermicidal agent	[16]
<i>Acacia catechu</i>	Fabaceae	Exudate	-	Effective oral contraceptive in rats and inhibits implantation	[15]
<i>Acacia concinna</i>	Fabaceae	Bark	Lupeol, α -spinosterol, hexacosanol, α -apinasterone, acacia acid, acacia acid lactone and an amorphous saponin	Spermicidal and semen coagulating activities. Saponin showed spermicidal activity.	[16,17, 18]
<i>Acalypha indica</i>	Euphorbiaceae	Plant	-	Post-coital anti-fertility activity	[19]
<i>Achrostichum aureum</i>	Pteridaceae	Plant	-	Anti-implantation activity	[20]
<i>Achyranthes aspera</i>	Amaranthaceae	Aerial part, Leaf	-	Anti-implantation activity	[21,22, 23]
<i>Adhatoda vasica</i>	Acanthaceae	Leaf	Vasicine	Anti-implantation activity	[24, 25]
<i>Adiantum capillus</i>	Adiantaceae	Plant	Isoadiantone	Inhibited post coital implantation in rats	[26]
<i>Allium cepa</i> L.	Liliaceae	Bulb	β -sitosterol, Kampferol	Abortifacient, Antifertility activity	[27]
<i>Albizia lebbek</i> (L.) Benth.	Leguminosae	Pod	Lebbekanin-E	Spermicidal activity	[28, 29]

<i>Albizzia procera</i>	Apocynaceae	Seed	Oleanolic acid saponin and Proceric acid saponin mixture	Spermicidal activity	[17]
<i>Alstonia scholaris</i>	Apocynaceae	Bark	Leupelol acetate	Antifertility effect in male rats.	[30, 31]
<i>Anagallis arvensis</i>	Primulaceae	Stem	-	Spermicidal and semen coagulating activities	[18]
<i>Ananas comosus</i> Mers.	Bromeliaceae	Leaf	Stigmastane, Sitosterol β , Ergosterol peroxide	Antiimplantation activity	[32]
<i>Ananas sativus</i>	Bromeliaceae	Fruit	-	Antiovolatory activity	[33]
<i>Andrographis paniculata</i>	Acanthaceae	Leaf, Stem	Andrographilo-de	Anti-spermatogenic and anti-androgenic effect	[34, 35]
<i>Androsace septentrionalis</i> L.	Primulaceae	Plant	Triterpene glycoside	Contraceptive and abortive action on rats and mice, prevented pregnancy in post-coital tests	[36, 37]
<i>Annona squamosa</i>	Annonaceae	Seed	-	Anti-implantation activity	[18]
<i>Aristolochia indica</i> L.	Aristolochiaceae	Root	Aristolic acid, p-coumaric acid, Methyl aristolate	Anti-spermatogenic and anti-androgenic effects	[38]
<i>Artabotrys odoratissimus</i>	Annonaceae	Leaf	-	Anti-implantation activity	[39]
<i>Asparagus pubescens</i>	Liliaceae/ Asparagaceae	Root	-	It inhibited fetal implantation.	[40]
<i>Austeroplenckia populnea</i>		Leaf	-	Decrease sperm concentration in cauda epididymides	[41]
<i>Azadirachta indica</i>	Meliaceae	Leaf, Seed	Neem oil	Anti-spermatogenic activities and histological changes in testes and epididymides. Prevented pregnancy if taken before sexual intercourse and	[42, 43, 44, 45, 46]

				checked embryo implantation.	
<i>Balanites roxburghii</i> L.	Zygophyllaceae	Fruit peel	Apigenin & Luteolin	Mass atrophy of spermatogenic elements due to secondary effects	[47]
<i>Bambusa vulgaris</i>	Poaceae	Spadix	-	Anti-ovulatory effect	[33]
<i>Barleria prionitis</i> L.	Acanthaceae	Root	-	Anti-fertility effect in male rats medicated by disturbances in testicular somatic cell functions.	[48]
<i>Beaumontia grandiflora</i>	Apocynaceae	Leaf	-	Showed anti-implantational, abortifacient and luteolytic effects	[49]
<i>Berberis chitria</i> Buch.-Ham ex. Lindl.	Berberidiaceae	Root	Palmitine Hydroxide	Impairment of germ cells	[50]
<i>Blepharispermu m subsessile</i>	Asteraceae	Rhizomes	-	Anti-implantation activity	[51]
<i>Bryocarpus coccineus</i> Schum	Connaraceae	Leaf	-	Increases spontaneous utrine muscle contraction	[52]
<i>Bursera fagaroides</i>	Burseraceae	Cortex	Glycosides	Human spermatozoa and those obtained from mouse epididymis became agglutinated and immobilized	[53]
<i>Butea frondosa</i>	Papilionaceae	Seed	-	Partial abortive in action in mice and rats	[54]
<i>Butea monosperma</i>	Papilionaceae	Seed	Butin	Potential male contraceptive with minimal side effects	[55]
<i>Caesalpinia decapetala</i>	Fabaceae	Aerial Part	-	Contraceptive activity	[56]
<i>Calotropis gigantea</i> L.	Asclpiadaceae	Root	Akundarin, Calotropin	Antiimplantation effect	[57]

<i>Calotropis procera</i> (Ait.) R.Br.	Asclepiadaceae	Root	Calotropin	Antispermatogenic and anti-implantation effects	[58]
<i>Carica papaya</i>	Caricaceae	Fruit, Seed	Papain & Pectin	Contraceptive effects	[59, 60, 61, 62]
<i>Casearia ilicifolia</i> Hochst.	lacourtiaceae	Leaf	Flavonoids, Triterpenes & steroids	Anti-fertility activity	[63]
<i>Casearia tomentosa</i>	Flacourtiaceae	Leaf	-	Interferes with spermatogenesis, anti-implantational and abortifacient	[49]
<i>Cassia fistula</i> L.	Fabaceae	Seed	Leucoanthocyanidin, Sennoside A & B, Rhein	Estrogenic activity	[64]
<i>Celastrus paniculatus</i>	Celastraceae	Seed	-	The testes of treated rats have shown vacuolisation, germ cell depletion arrest of spermatogenesis	[65]
<i>Centella asiatica</i>	Umbelliferae	Plant	Isothankuniside and BK Compound [Methyl-5-hydroxide-3,6-diketo-23(or 24)-norurs-12-en-28-oate]	Used as oral antifertility and both compounds caused consistent reduction of fertility	[66]
<i>Chordia dichotoma</i>	Boraginaceae	Leaf	-	Anti-implantational and abortifacient effects	[49]
<i>Cichorium intybus</i>	Asteraceae	Seed	-	Showed significant contraceptive activity	[67]
<i>Citrullus colocynthis</i>	Cucurbitaceae	Fruit	-	Anti-androgenic activity	[68]
<i>Citrus hystrix</i>	Rutaceae	Fruit peel	-	Estrogenic effect	[69]
<i>Citrus limon</i>	Rutaceae	Seed	-	Decreases the sperm motility and sperm count	[70]

<i>Coccus lacca</i>	Lacciferidae	Bark	-	Anti-implantation effect	[71]
<i>Codonospis ovate</i>	Campanulaceae	Plant	-	Anti-implantation activity	[24]
<i>Colebrookia oppositifolia</i>	Lamiaceae	Leaf	-	Effects on testicular cell population	[72]
<i>Coleus barbatus</i>	Labiatae	Leaf	-	Pregnant rats on treatment with extract before embryo implantation caused delayed fetal development	[73]
<i>Combretodendron africanum</i>	Lecythidaceae	Stem bark	Tannins & Saponisides	Caused abortion	[74]
<i>Corchorus olitorius</i>	Tiliaceae	Seed	-	Increases the carbonic anhydrase activity in the uterus of mice and elevates the level of progesterone	[75]
<i>Crotalaria juncea</i>	Fabaceae	Seed	-	Arrest spermatogenesis and is likely to have anti-androgenic activity	[76]
<i>Croton roxburghii</i>	Euphorbiaceae	Bark	-	Anti-steroidogenic activity	[77]
<i>Curcuma longa</i>	Zingiberaceae	Tuber	-	Spermatogenesis action	[78]
<i>Cuscuta reflexa</i>	Convolvulaceae	Stem	-	Increases the carbonic anhydrase activity in the uterus of mice and elevates the level of progesterone	[79]
<i>Cynomorium coccineum</i>	Cynomoriaceae	Stem, Root	-	Effect on epididymal sperm pattern	[80]
<i>Daphne species</i>	Thymellaeaceae	Plant	Yuahautin	Contraceptive and abortifacient effects	[81]
<i>Daucus carota</i>	Apiaceae	Seed	Volatile oil, carbohydrate fraction of Carrot seed oil	Estrogenic effect	[82]
<i>Derris brevipes</i>	Asteraceae	Root	-	Anti-implantation activity	[83]
<i>Deutzia corymbosa</i>	Hydrangeaceae	Plant	-	Antifertility activity	[84]

<i>Dictamnus albus</i>	Rutaceae	Root bark	Fraxinellone	Antifertility activity	[85]
<i>Dieffenbachia amoena</i>	Araceae	Leaf	-	Antifertility activity	[86]
<i>Diospyros embryopteris</i>	Ebenaceae	Leaf	-	Reduces sperm motility	[49]
<i>Diploclisia glaucescens</i>	Menispermaceae	Stem	Ecdysterone	Spermicidal activity	[87]
<i>Dipsacus mitis</i>	Dipsacaceae	Root	-	Antifertility activity	[88]
<i>Dysoxylum binectariferum</i>	Meliaceae	Stem bark	Rohitukene	Contraceptive agent	[89]
<i>Echeveria gibbiflor</i>	Crassulaceae	Leaf	-	Contraceptive agent	[90]
<i>Echinops echinatus</i>	Asteraceae	Root	-	Sperm anti-motility	[91]
<i>Embelia ribes</i> Burm.f.	Myrsinaceae	Seed, Root, Bark	Embelin	Potent oral contraceptive, antifertility activity	[92, 93]
<i>Ensete superpa</i>	Musaceae	Seed	Kadalin	Antifertility activity	[94]
<i>Ephedera gerardiana</i>	Ephedraceae	Plant	-	Antifertility activity	[84]
<i>Epilobium angustifolium</i>	onagraceae	Plant	-	Antifertility activity	[95]
<i>Eugenia jambolana</i>	Myrtaceae	Flower	Oleanolic acid	Spermatogenic activity	[96]
<i>Ferula foetida</i>	Umbellifera	Rhizomes	-	Anti-implantation activity	[97]
<i>Ferula jaesochkeana</i>	Umbellifera	Aerial part	Ferujol	Contraceptive agent	[98, 99]
<i>Foeniculum vulgare</i>	Apiaceae	Seed	Anethole	Anti-implantation activity	[100]

<i>Gardenia jasminoides</i>	Rubiaceae	Flower	Cycloartane triterpenoids namely gardenic acid and gardenolic acid B	Terminated early pregnancy in rats	[101]
<i>Geranium lucidum</i>	Geraniaceae	Plant	-	Antifertility activity	[84]
<i>Gleditschia horrid</i>	Fabaceae	Pod	Saponins	Anti-implantation activity	[102]
<i>Globularia alypum</i>	Globulariaceae	Leaf	-	Anti-implantation activity	[103]
<i>Globularia Arabica</i>	Globulariaceae	Leaf	-	Anti-implantation activity	[103]
<i>Gloriosa superba</i> L.	Liliaceae	Root	Colchicine	Antiimplantation activity	[104]
<i>Gnaphalium indicum</i>	Compositae	Plant	-	Antifertility activity	[84]
<i>Gossypium barbadense</i>	Malvaceae	Seed	Gossypol	Antifertility activity	[105]
<i>Gossypium herbaceum</i>	Malvaceae	Seed	Gossypol acetic acid	Contraceptive effect	[106, 107]
<i>Guaiacum officinale</i>	Zygophyllaceae	Aerial part	Saponins	Abortifacient activity	[108]
<i>Guetterda andamanica</i>	Meliaceae	Aerial part	-	Contraceptive activity	[67]
<i>Heliotropium indicum</i>	Boraginaceae	Plant	n-hexacosanol, Sitosterol, Stigmasterol, Chalinasterol & Campesterol	Antifertility activity	[109]
<i>Hibiscus macranthus</i>	Malvaceae	Leaf	-	Effect on testicular function	[110]

<i>Hibiscus rosa-sinensis</i>	Malvaceae	Flower, Leaf	-	Anti-spermatogeni, antiandrogenic, Antifertility, anti-implantation activities	[111, 112, 113, 114]
<i>Hyptis suaveoleus</i>	Labiatae	Plant	-	Antifertility activity	[115]
<i>Ixora finlaysoniana</i>	Rubiaceae	Aerial part	-	Antifertility activity	[116]
<i>Juniperus communis</i>	Cupressaceae	Seed	-	Inhibit pregnancy	[24]
<i>Kigelia pinnata</i>	Bignoniaceae	Plant	-	Anti-implantation activity	[24]
<i>Leonotis ocyimifolia</i>	Lamiaceae	Leaf, Root	Leonitin	Antifertility & Abortifacient effect	[117]
<i>Lepidium capitatum</i>	Cruciferae	Plant	-	Anti-implantation activity	[24, 118]
<i>Leucas cephalotes</i> (Roth.)	Lamiaceae	Fruit	Oleanolic acid, 7-oxositosterols, stigmasterol, β - sitosterols	Antifertility activity	[119]
<i>Lindenbergia indica</i>	Scrophulariaceae	Plant		Arrests oogenesis	[120]
<i>Lygodium flexuosum</i> L.	Schizaeaceae	Plant	-	Anti-ovulatory and anti-implantation activity	[121]
<i>Malvaviscus konzattii</i>	Malvaceae	Flower	-	Contraceptive effect	[122]
<i>Marsdenia koi</i>	Apocynaceae	Plant	Steroidal glycosides: Marsdekoiside A & B	Antifertility activity	[123]
<i>Marsilea minuta</i>	Marsileaceae	Plant	-	Elevates the level of cholesterol and ascorbic acid content of ovaries	[124]
<i>Martynia annua</i>	Martyniaceae	Root	-	Suppresses testicular and epididymal sperm counts and caused lesions on seminiferous tubules	[125]

<i>Maytenus ilicifolia</i>	Celastraceae	Leaf	-	Antifertility activity	[126]
<i>Memecylon lushingtonii</i>	Meliaceae	Aerial part	-	Contraceptive activity	[67]
<i>Mentha arvensis</i>	Lamiaceae	Leaf	-	Antifertility activity	[127]
<i>Milletia auriculata</i>	Fabaceae	Leaf	-	Reduces sperm motility	[49]
<i>Momordica charantia</i>	Cucurbitaceae	Seed	β –momorcharin & Sterols	Anti-spermatogenic effect	[128, 129]
<i>Momordica cochinchinesis</i>	Cucurbitaceae	Root, Tuber	Momorcochin	Abortifacient activity	[130]
<i>Momordica dioica</i>	Cucurbitaceae	Tuber	-	Spermicidal activity	[131]
<i>Mondia whietii</i>	Periplocaceae	Root	-	Inhibitory effects on spermatogenesis and reduces fertility	[132]
<i>Montanoa tomentosa</i>	Asteraceae	Leaf	Kaurenoic acid, Kauredionioic acid	Anti-fertility activity	[133]
<i>Moringa oleifera</i>	Moringaceae	Root	-	Anti-progestational and antifertility activity	[134]
<i>Mucuna urens</i>	Faboideae/ Leguminosae	Seed	-	Arrests spermatogenesis at spermatid stage	[135]
<i>Murraya paniculata</i>	Rutaceae	Root	Yuehchukene	Anti-implantation activity	[136, 137]
<i>Nelumbo nucifera</i> Gaerth.	Nymphaeaceae	Seed	-	Anti-progestational and anti-estrogenic activities	[138]
<i>Nicotiana tabaccum</i>	Solanaceae	Aerial part	Nicotine	Anti-androgenic effect	[139]
<i>Nigella sativa</i> L.	Ranunculaceae	Seed	-	Antifertility activity	[140]

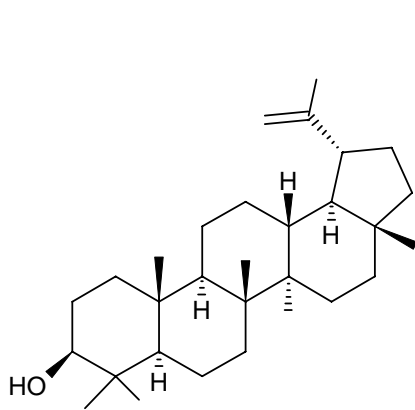
<i>Ocimum sanctum</i> L.	Labiatae/ Laminaceae	Leaf	-	Antifertility activity	[141]
<i>Pentapanax leschenaultii</i>	Araliaceae	Fruit	Oleanolic acid, Triterpene glycosides	Spermicidal activity	[142]
<i>Phonix dactilifera</i>	Arecaceae	Fruit	-	Anti-ovulatory activity	[33]
<i>Phyllanthus amarus</i>	Phyllanthaceae	Plant	-	Anti-implantation activity	[143]
<i>Physalis minima</i> L.	Solanaceae	Plant	-	Anti-implantation activity	[144]
<i>Phytolacca americana</i>	Phytolaccaceae	Root	Steroidal acidic saponin	Estrogenic effects	[145]
<i>Phytolacca dodecandra</i>	Phytolaccaceae	Bark	Saponins, Lemmatoxin, Oleanoglycotoxin-A, Lemmatoxin-C	Antifertility activity	[146]
<i>Piper longum</i> L.	Piperaceae	Fruit	Piperine	Anti- spermetogenic effect	[93]
<i>Pisum sativum</i>	Fabaceae	Seed	m-Xylohydroquinone	Oral contraceptives inhibits endometrial development in rats	[147, 148]
<i>Pithecolobium saman</i>	Fabaceae	Flower	Samanin-D	Spermicidal activity	[149]
<i>Plantanus orientalis</i>	Platanaceae	Bark, Leaf	-	Anti-implantation activity	[84]
<i>Plumbago zeylanica</i>	Plumbaginaceae	Root	Plumbagin	Anti-implantation , abortifacient activity in rats	[150, 151]
<i>Punica grantum</i>	Punpacaceae	Plant without root	-	Anti-implantation activity	[84]
<i>Quasia amara</i>	Simaroubaceae	Bark	-	Anti- spermatogenic effect	[152]

<i>Randia dumetorum</i>	Rubiaceae	Seed	Oleanolic acid glycoside	Anti-implantation activity	[153]
<i>Ricinus communis</i> L.	Euphorbiaceae	Seed	-	Showed anti-implantation activity	[154]
<i>Rivea hypocrateriformis</i>	Convolvulaceae	Aerial part	-	Anti-implantation activity	[155]
<i>Rubus ellipticus</i>	Asteraceae	Plant without root	-	Anti-implantation activity	[24]
<i>Ruellia prostrate</i>	Acanthaceae	Plant	Lupeol, Sitosterol, Stigmasterol & long chain ester (C ₄₂ -C ₆₀)	Anti-fertility activity	[156]
<i>Rumex steudelli</i>	Polygonaceae	Root		Prolonged diestrous stage in rats. Produced anti-fertility effect.	[157, 158]
<i>Ruta gravelans</i> L.	Rutaceae	Root, Stem, Leaf	Chalepensisin	Showed significant anti-fertility activity in rats at early stages of pregnancy	[159]
<i>Salsola tuberculatiformis</i>	Chenopodiaceae	Plant	2-(4-acetoxyphenyl)-2-chloro-N-methyl-ethyl ammonium chloride (Compound A)	Contraceptive effect on female rats.	[160]
<i>Salvia fruticosa</i>	Lamiaceae	Leaf	-	Viable fetuses in females	[161]
<i>Saraca indica</i>	Fabaceae	Bark	-	Showed anti-progestational activity in rabbits. Exhibited anti-implantation effect	[71]
<i>Sarcostemma acidum</i>	Asclepidaceae	Stem	-	Arrests spermatogenesis in male rats without noticeable side effects	[162]
<i>Schefflera capitata</i>	Araliaceae	Plant	Schefferoside	Spermicidal activity	[163]

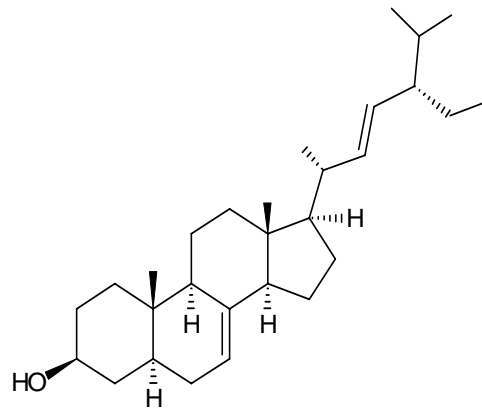
<i>Senecio vulgaris</i>	Asteraceae	Plant	Senicionine & Senicionine-N-oxide	Decreased the number of normal fetuses per pregnant rat	[164]
<i>Sesbania sesban</i>	Fabaceae	Seed	oleanolic acid 3-beta-d-glucuronide	Anti-implantation activity	[165]
<i>Solanum crassypetalum</i>	Solanaceae	Aerial part	-	Showed significant contraceptive activity in adult female rats	[67]
<i>Sponias mombin</i>	Anacardiaceae	Leaf	-	Induces abortion	[166]
<i>Stephania hernandifolia</i>	Menispermaceae	Leaf	Steroid	Spermicidal activity	[22]
<i>Stevia rebaudiana</i>	Astraceae	Plant	-	Decrease in testosterone level	[167]
<i>Striga lutea</i> Lour.	Scrophulariaceae	Plant	Acacetin, Luteolin & Flavones	Possesses significant anti-fertility activity in mice. Exhibits estrogenic property	[168]
<i>Striga orobanchioides</i>	Scrophulariaceae	Plant	Apigenin & Luteolin	Exhibited slight anti-estrogenic activity. Dose- dependent anti-implantation activity	[169]
<i>Strumpfia maritime</i>	Rubiaceae	Flowering top	-	Anti-fertility activity	[170]
<i>Taxus baccata</i> L.	Taxaceae	Leaf	-	Inhibited pregnancy in 60% of albino rats	[171]
<i>Tephrosia purpurea</i> L.	Fabaceae	Seed	Purpurin, Rutin	Anti-fertility activity	[172]
<i>Tinospora cordifolia</i>	Menispermaceae	Stem	-	Decreases sperm motility as well as sperm density, protein, glycogen contents of the testis	[173]
<i>Toddalia asiatica</i> (L.) Lam.	Rutaceae	-	Berberine, Toddaline, Toddalinine	Abotifacient	[174]
<i>Trichosanthes</i>	Cucurbitaceae	Root,	β - Trichosanthin	Induced mid-term abortion in mice and	[175]

<i>cucumeroides</i>		Tuber		twice as potent as trichosanthin	
<i>Trichosanthes kirilowi</i>	Cucurbitaceae	Root, Tuber	Trichosanthin	Induced mid-term abortion	[175]
<i>Trigonella foenum-graceum</i> L.	Papilionaceae/Fabaceae	Seed	Diosgenin, Yamogenin and Tigogenin	Exerts both anti-fertility and antiandrogenic activity.	[176, 177]
<i>Tripterygium wilfordii</i> Hook.	Celastraceae	Root	Total glucosides, Diterpene epoxides, Triptolide, triptidiolide, Triptolideenol,	Safe, reversible male anti-fertility agent. Anti-apermatogenic activity similar to that of gossypol. induce complete infertility male rats	[178, 179]
<i>Vicoa indica</i> L.	Asteraceae	Plant	Vicolid B, Vicolid D	Anti-implantation activity	[180]
<i>Vigna unguiculata</i> L.	Fabaceae	Pod	-	Reduces spermatogenesis	[181]
<i>Vinca rosea</i>	Apocynaceae	Aerial part	Vincristine	Decrease in secretory activity of sex glands. Epididymal dysfunction.	[182]
<i>Vitex negundo</i> L.	Verbenaceae	Seed	Flavonoid-rich fraction (5,7,3'-trihydroxy-6,8,4'-trimethoxy flavones)	anti-implantation effects	[183]
<i>Ziziphus jujube</i>	Rhamnaceae	Bark	-	Reversible anti-steroidogenic activity	[77]

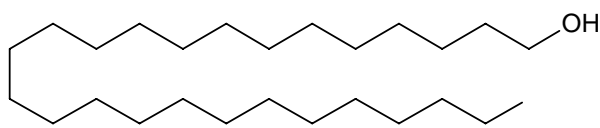
Figure: 1 Structure of some chemical constituents isolated from listed plants



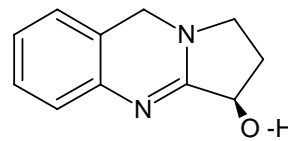
Lupeol



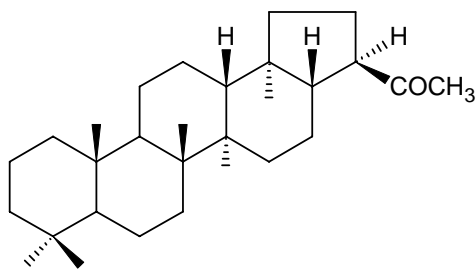
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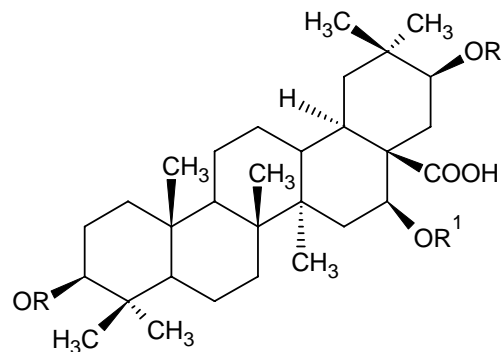
Hexacosanal



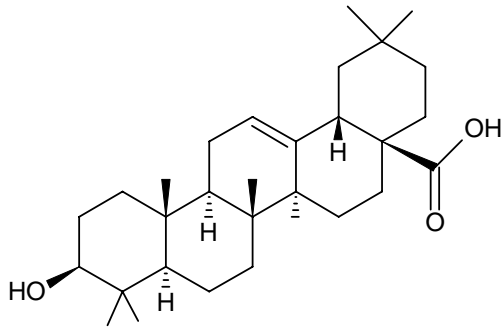
Vasicine



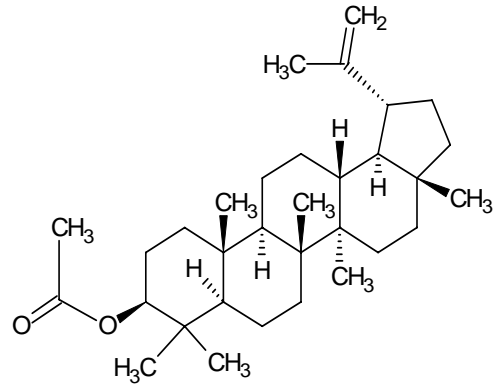
Isodiantone



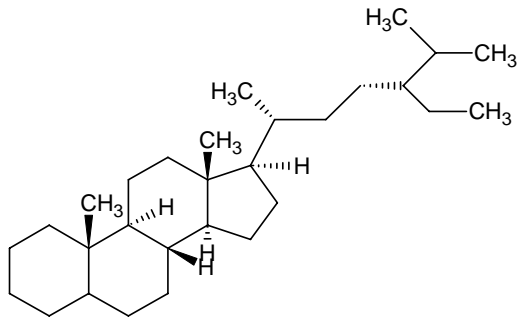
Lebbekinin -E



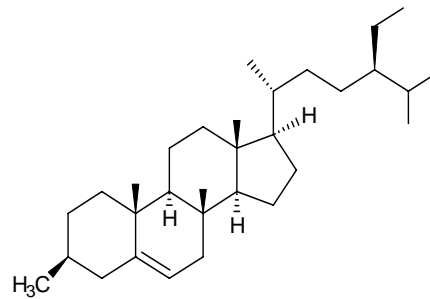
Oleanolic acid



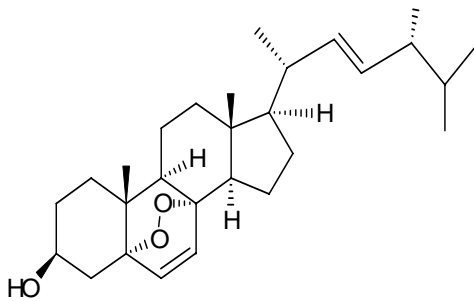
Leupelol acetate



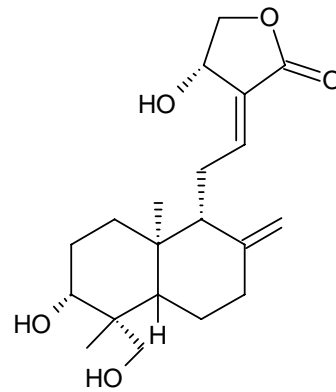
Stigmastane



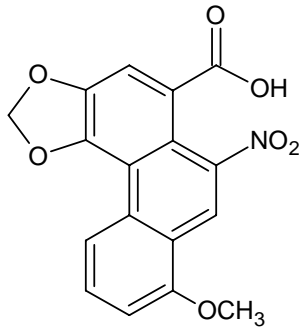
Beta - Sitosterol



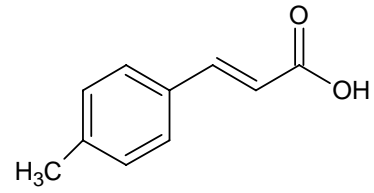
Ergosterol peroxide



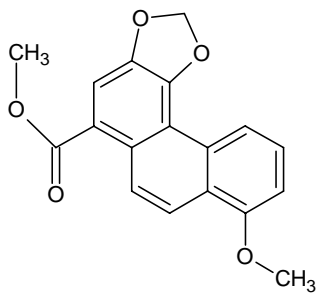
Andrographolide



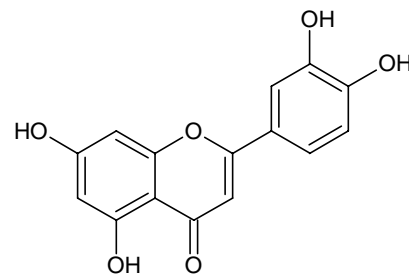
Aristolic acid



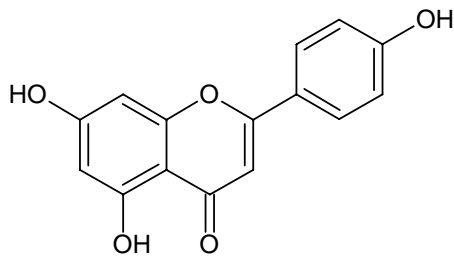
p -coumaric acid



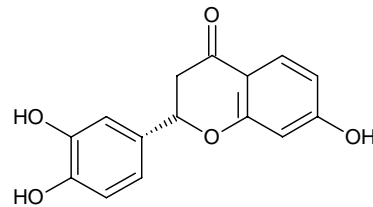
Methyl aristolate



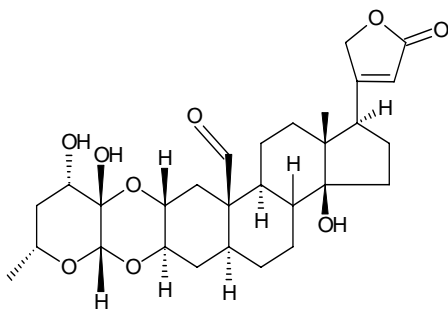
Luteolin



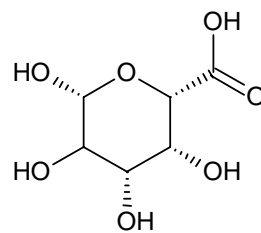
Apigenin



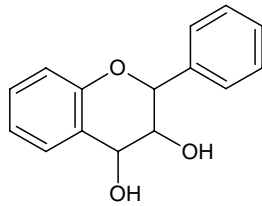
Butin



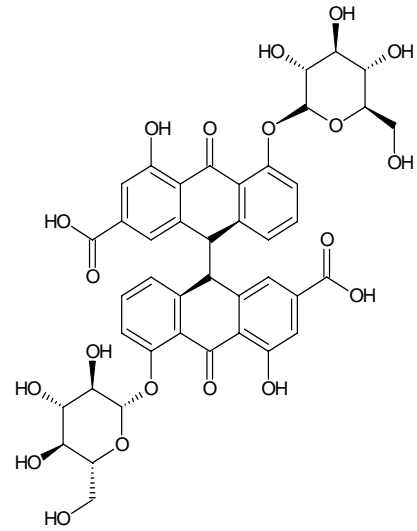
Calotropin



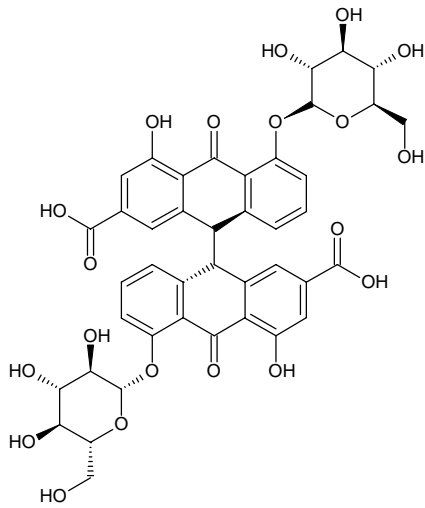
Pectin



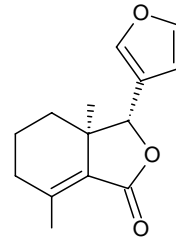
Leucoanthocyanidin



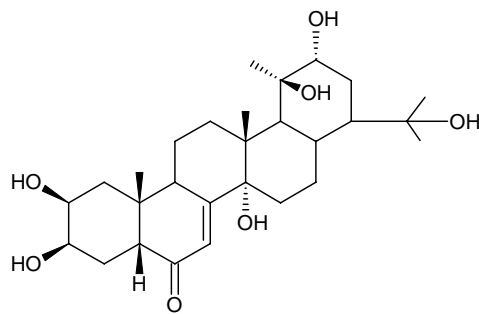
Sennoside A



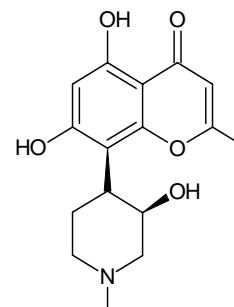
Sennoside B



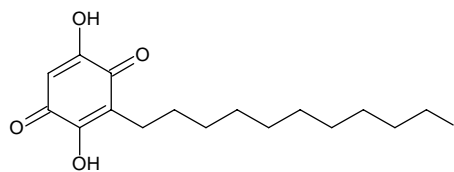
Fraxinellone



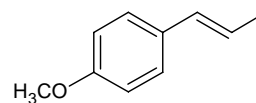
Ecdysterone



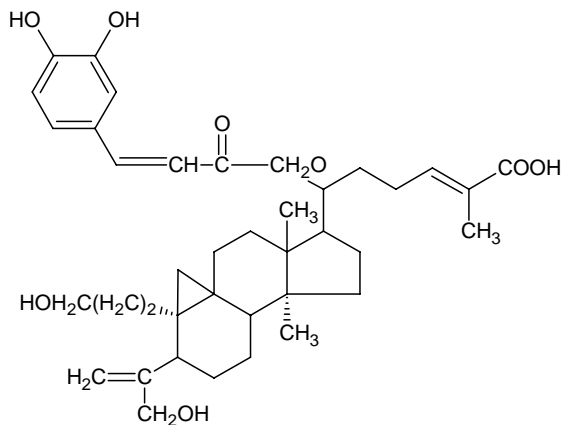
Rohitukene



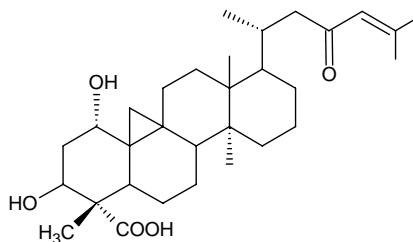
Embelin



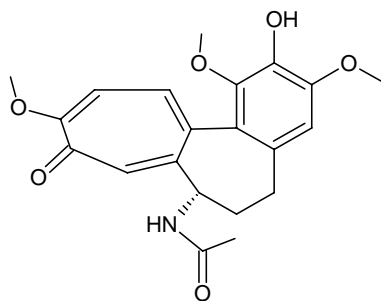
Anethole



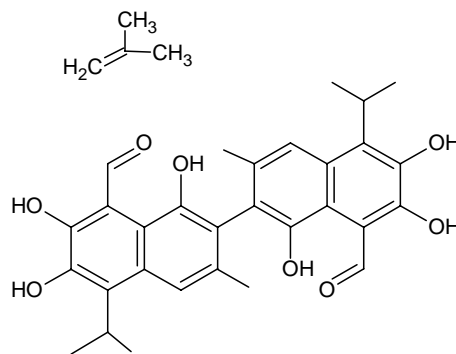
Gardenic acid



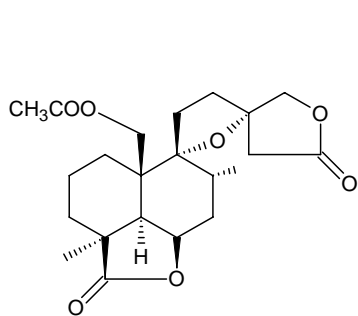
Gardenolic acid B



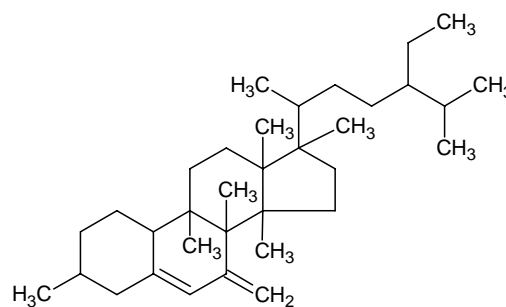
Colchicine



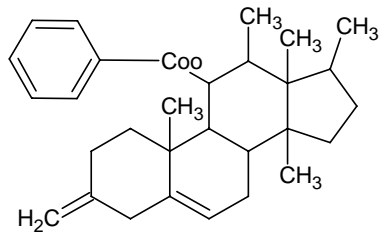
Gossypol acetic acid



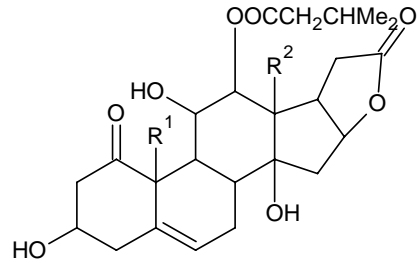
Leonitin



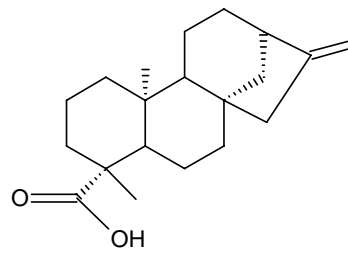
7-Oxo-beta-sitosterol



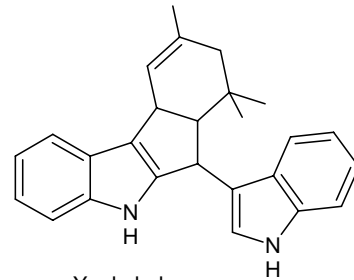
Tinctoramine



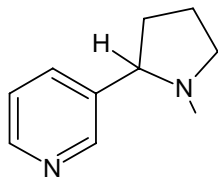
Tinctoralactone



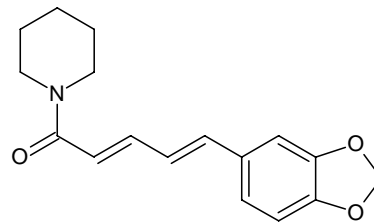
kaurenoic acid



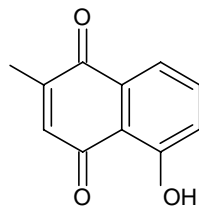
Yuehchukene



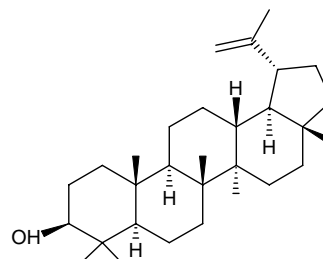
Nicotine



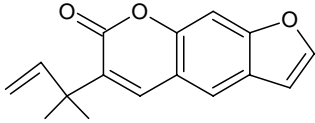
Piperine



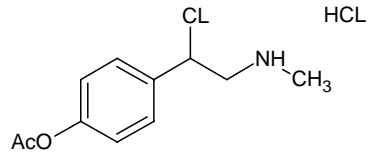
Plumbagin



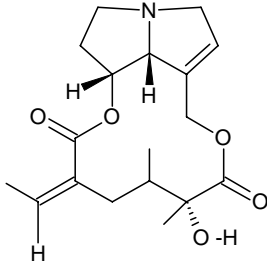
Lupeol



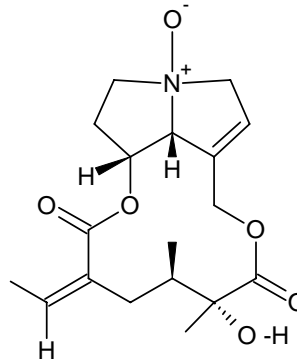
Chalepensin



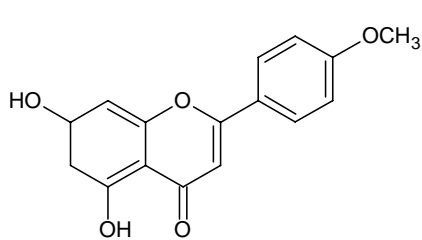
2-(4-acetoxophenyl)-2-chloro-N-methyl-ethyl ammonium chloride



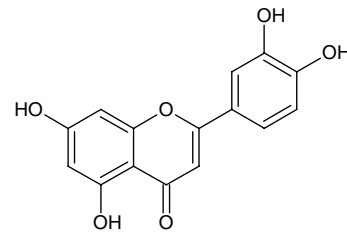
Senecionine



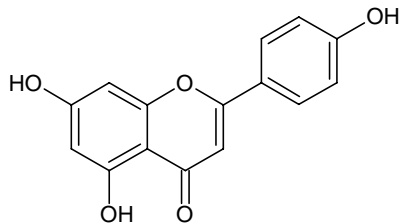
Senecionine N-oxide



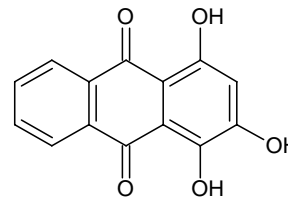
Acacetin



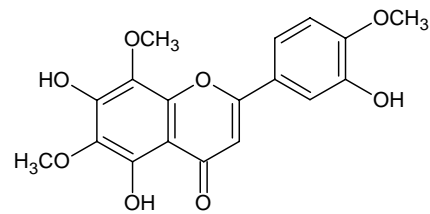
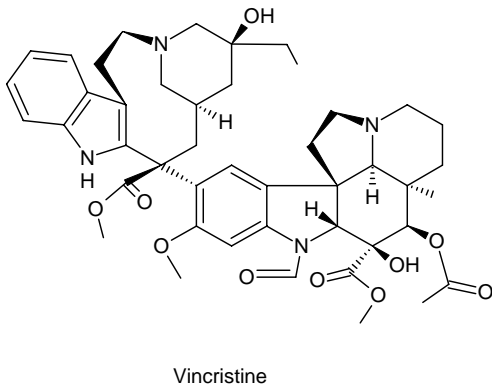
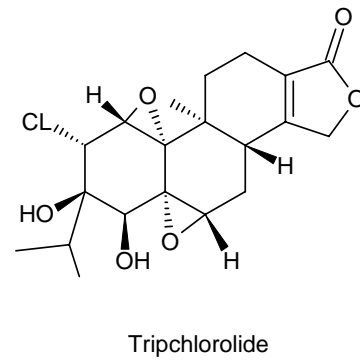
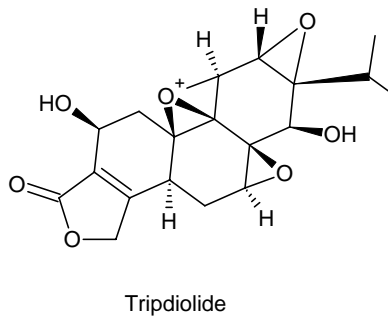
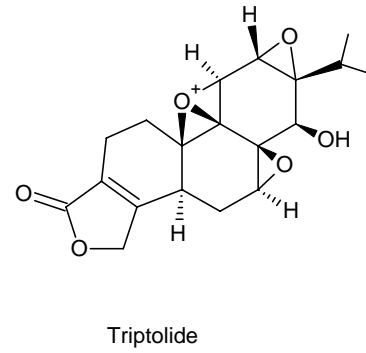
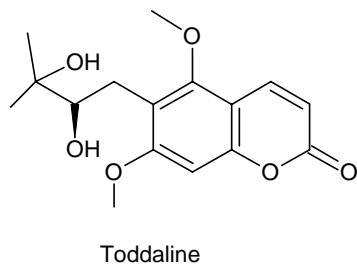
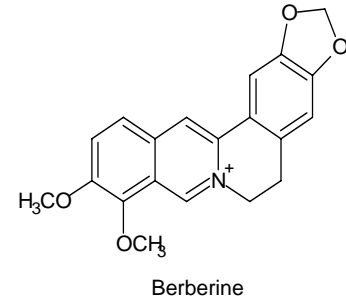
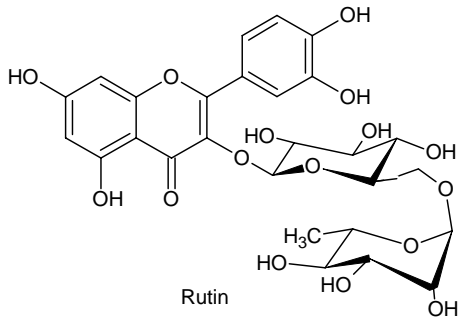
Luteolin

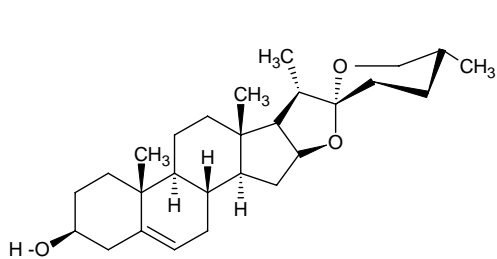


Apigenin

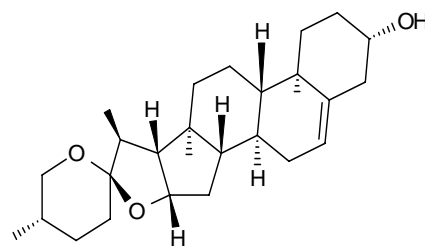


Purpurin

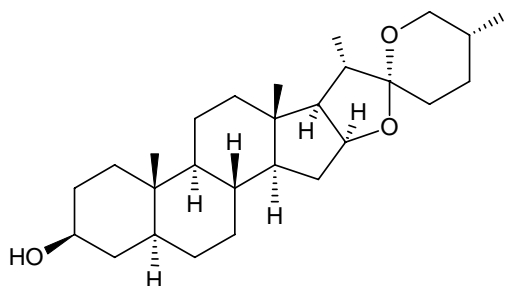




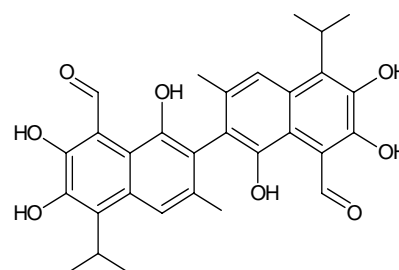
Diosgenin



Yamogenin

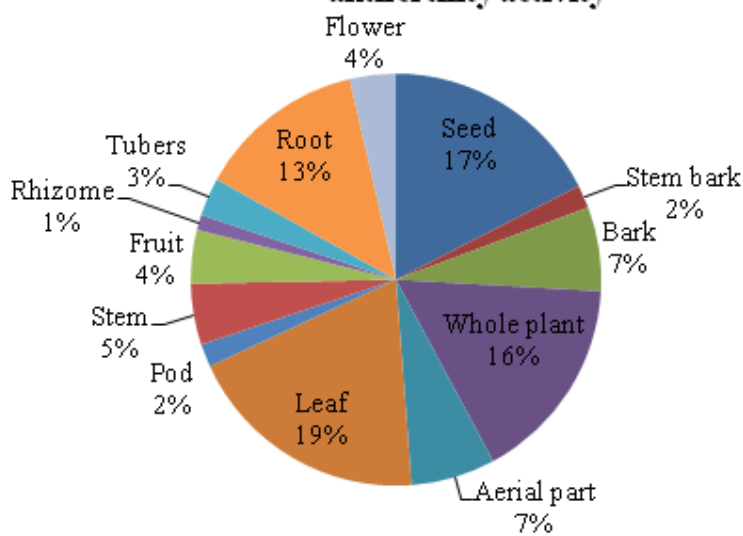


Tigogenin



Gossypol

Figure: 2 Percentage of Different plant parts responsible for antifertility activity



REFERENCES

1. Chaudhary A., Sharma N., Sharma P., Jasuja N.D., Sharma G., Contraceptive property of Nitrogen/oxygen sulphur donor hetrocyclic compounds. *Rasayan Journal of Chemistry* 2008; 3: 648-692.
2. Sharma J.D., Sharma L., Yadav P., Antifertility Efficacy of *Piper* betle Linn. (Petiole) on Female Albino Rats. *Asian J Exp Sci*, 2007; 21(1): 145-150.
3. <http://www.overpopulation.org/solutions.html>
4. <http://www.ibiblio.org/lunarbin/worldpop-population world>
5. Sharma R.S., Rajalakshmi M., Antony Jeyaraj D., Current status of fertility control methods in India. *J Biomed Sci*, 2001; 26(4): 391-405.
6. James S., Kepron C., Lemons, yams and crocodile dung: a brief history of birth control. *U Toronto Med J*, 2002; 79(1): 156-158.
7. Tripathi K.D., *Essentials of Medical Pharmacology*, Jaypee Medical Publishers, Kolkata, 6th ed. 2008.
8. Rostum A.H., Makonnen E., Zerihun L., Assefa G., Toxicity stuy of *Jatropha curcas* and *Ricinus communis* seed extracts in experimental animals. *Egypt J Med Lab Sci*, 1998; 7(2): 93-101.
9. Singh M.M., Effect of some non-steroidal anti-fertility agents on histology and biochemistry of the uterus and uterine fluid during delayed implantation in rats. *Contraception*, 1973; 8: 235-253.
10. Sandhya B., Thomas S., Isabel W., Ethnomedicinal plants used by the Valaiyan community of Piranmalai Hills (Reserved Forest), Tamil Nadu, India. A pilot study. *Afr.J. Traditional Complementary Altern Med*, 2006; 3(1): 101-114.
11. Adailkan P.G., Gauthaman K., The aging male, 2001; 4: 163-169.
12. Zia-UI-Haque A., Quazi M.H., Hamdard M.E., Studies on the antifertility properties of active components isolated from seeds of *Abrus precatorius* Linn. I. *Pakistan J Zool*, 1983a; 15(2): 129-139.
13. Zia-UI-Haque A., Quazi M.H., Hamdard M.E., Studies on the antifertility properties of active components from seeds of *Abrus precatorius* Linn. II. *Pakistan J Zool*, 1983b; 15(2): 141-146.
14. Sinha S., Mathur R.S., Effect of steroidal fraction of seeds of *Abrus precatorius* Linn. On rat testis. *Indian J Exp Biol*, 1990; 28(8): 752-756.
15. Choudhary A.K.A., Chakder S.K., Antifertility activity of Traditional contraceptive pill comprising *Acacia catechu*, *Acacia Arabica* and *Traquia involucrata*. *Indian J Med Res*, 1984; 80: 372-374.
16. Banerji R., Nigam S.K., Chemistry of *Acacia cocinna* anmd *A. caesia* bark. *J Indian Chem Soc*, 1980; 57(10): 1043-1044.
17. Banerji R., Srivastava A.K., Misra G., Nigam S.K., Steroid and triterpenoid saponins as spermicidal agents. *Indian Drugs*, 1979; 17(1): 6-8.
18. Kamboj V.P., Dhawan B.N., Research on plants for fertility regulation in India. *J Ethnopharmacol*, 1982; 6: 191-226.
19. Hiremath S.P., Rudresh K., Badami S., Patil S.B., Post-coital antifertility activity of *Acalypha indica* L. *J Ethnopharmacol*, 1999; 67: 253-257.
20. Dhar J.D., Setty B.S., Lakshmi V., Bhakuni D.S., Post- coital antifertility activity of the marine plant, *Achrosticum aureum* L. in rats. *Indian J Med Res*, 1992; 96: 150-152.

21. Wadhwa V., Singh M.M., Gupta D.N., Singh C., Contraceptive and hormonal properties of *Achyranthes aspera* in rats and hamsters. *Planta Med*, 1986; 3: 231-233.
22. Paul D., Bera S., In vitro determination of the contraceptive spermicidal activity of a composite extract of *Achyranthes aspera* and *Stephania hernandifolia* on human semen. *Contraception*, 2006; 73: 284-288.
23. Shibeshi W., Makonnen E., Zerihun L., Effect of *Achyranthes aspera* L. on fetal abortion and pituitary weights, serum lipids and hormones. *Afr Health Science*, 2006; 6: 108-112.
24. Prakash A.O., Saxena V., Shukla S., Mathur S., Gupta A., Sharma S., Anti-implantation activity of some indigenous plants in rats. *Acta Europaea Fertilitatis*, 1985; 16(6): 441-448.
25. Rachana, Basu S., Pant M., Kumar M.P., Saluja S., Review & Future Perspectives of Using Vasicine, and Related Compounds. *Indo Global J Pharm Sci*, 2011; 1(1): 85-98.
26. Murthy R.S.R., Antiimplantation activity of isoadiantone. *Indian Drugs*, 1984; 21(4): 141-144.
27. Thakare V.N., Kothavade P.S., Dhote V.V., Deshpande A.D., Antifertility Activity of Ethanolic Extract of *Allium cepa* Linn in Rats. *Int J Pharm Tech Res*, 2009; 1(1): 73-78.
28. Varshney I.P., Vyas P., Srivastava H.C., Singh P.P., Study of *Albizzia lebbek* Benth wood seaponin. *Lebbekanin-E. Nat Acad Sci Lett (India)*, 1979; 2(4): 135-136.
29. Gupta R.S., Kachhawa J.B., Chaudhary R., Antifertility effects of methanolic pod extract of *Albizzia lebbek* (L.) Benth. In male rats. *Asian J Androl*, 2004a; 6(2): 155-159.
30. Gupta R.S., Sharma R., Sharma A., Bhatnagar A.K., Dobhal M.P., Effect of *Alostonia scholaris* bark extract on testicular function of Wistar rats. *Asian J Androl*, 2002b; 4(3): 175-178.
31. Gupta R.S., Bhatnagar A.K., Joshi Y.C., Sharma M.C., Induction of antifertility with leupelol acetate in male albino rats. *Pharmacology*, 2005; 75(2): 57-62.
32. Pakrashi A., Basak B., Abortifacient effect of steroids from *Ananas comosus* and their analogues on mice. *J Reprod Fert*, 1976; 4: 461-462.
33. Bhaskaran S., Khanna S., Effect of alcoholic extract of *Ananas bamboosa* and *Phoenix oestrus* cycle of female albino rat. *Indian J Nat Prod*, 2001; 17(2): 43-46.
34. Gupta R.S., Sharma R., A review on medicinal plants exhibiting antifertility activity in males. *Natural Product Radiance*, 2006; 5(5): 389-410.
35. Akbarsha M.P., Manivanna B., Hamid K.S., Antifertility effect of *Andrographis paniculata* (Nees) in male albino rats. *Indian J Exp Biol*, 1990; 28: 421-426.
36. Mats M.N., Khorkov V.V., Kransnov E.A., Pirozhkova N.M., Chemical composition and contraceptive activity of *Androsace septentrionalis* L. *Rastit Resur*, 1984; 20(3): 403-408.
37. Mats M.N., Savchenko O.N., On the mechanism of the contraceptive effect of triterpene glycosides of *Androsace septentrionalis* L. *Farmakol Toksikol*, 1986; 49(2): 38-39.
38. Che C.T., Ahmed M.S., Kang S.S., Waller D.P., Studies on *Aristolochia III*-Isolation and Biological evaluation of *Aristolochia indica* roots for fertility regulation activity. *Nat Prod*, 1984; 47(2): 331-341.
39. Hegde D.A., Khosda R.L., Chansouria J.P.N., Sahal M., *Artabotrys odoratissimus*-A possible antifertility agent. *Indian J Nat Prod* 1993; 9(1): 15-16.
40. Nwafor P.A., Okwuasaba F.K., Onoruvwe O.O., Contraceptive and non-estrogenic effects of methanolic extract of *Asparagus pubescens* root experimental animals. *J Ethnopharmacol* 1998; 62(2): 117-122.

41. Mazaro R., De Stasi L., De Grava Kempinas W., Effects of hydromethanolic extract of *Austroplenckia populena* (Celastraceae) on reproductive parameters of male rat. *Contraception* 2002; 66(3): 205-209.
42. Sinha K.C., Riar S.S., Tiwary R.S., Dhawan A.K., Bardhan J., Neem oil as a vaginal contraceptive. *Indian J Med Res*, 1984; 79: 131-136.
43. Tewari R.K., Mathur R., Prakash A.O., Post-coital antifertility effect of neem oil in female albino rats. *IRCS Med Sci*, 1986; 14(10): 1005-1006.
44. Riar S.S., Bardhan J., Thomas P., Kain A.K., Mechanism of antifertility action of neem oil. *Indian J Med Res*, 1988; 88: 339-342.
45. Shaikh P.D., Manivannan B., Pathan K.M., Antispermatic activity of *Azadirachta indica* leaves in albino rats. *Current Science*, 1993; 64: 688-689.
46. Upadhyay S.N., Antifertility effect of neem (*Azadirachta indica*) oil in male rats by single intra-vas administration an alternate approach to vasectomy. *J Androl*, 1993; 14(4): 275-281.
47. Dixit V.P., Hbargava S.K., Gupta R.A., Hyperglycemia induced testicular dysfunction after chronic administration of *Balanites roxburghii* Planch fruit pulp extract in dogs *Canis indicus* L. *Indian J Exp Biol*, 1981; 19(10): 918-921.
48. Gupta R.S., Kumar P., Dixit V.P., Dobhal M.P., Antifertility studies of the root extract of *Barleria prionitis* Linn. In male albino albino rats with special reference to testicular cell population dynamics. *J Ethnopharmacol*, 2000; 70: 111-117.
49. Choudhary D.N., Singh J.N., Verma S.K., Singh B.P., Antifertility effects of leaf extracts of some plants in male rats. *Indian J Exp Bio*, 1990; 28: 714-716.
50. Gupta R.S., Dixit V.P., Testicular cell population following dynamics palmitine hydroxide treatment in dogs. *J Ethnopharmacol*, 1989; 25:151-157.
51. Agarwal S.K., Verma S., Singh S.S., Kumar S., Keshri G., Antifertility chromene from *Blepharispermum sussesile*. *Fitoterapia*, 1999; 70: 435-437.
52. Amos S., Binda L., Kunle O.F., Wambebe C., Uterotonic properties of ethanol extract of *Bryocarpus cocineus*. *Pharm Biol*, 2002; 40(1): 33-38.
53. Huacuja R.L., Delgado N.M., Carranco L.A., Reyes L.R., Rosado G.A., Agglutinating and immobilizing activity of an ethanol extract of *Bursera fagaroides* on human and mammalian spermatozoa. *Arch Invest Med (Mex.)*, 1990; 21(4): 393-398.
54. Kapila K., Bhide N.K., Razdan M.K., The antifertility effect of *Butea frondosa* petal (alcoholic extract and its crystalline fraction). *J Indian Med Assoc*, 1970; 1655(2): 60-61.
55. Bhagrava S.K., Estrogenic and Post coital anticonceptive activity in rats of butin b isolated from *Butea monosperma* seed. *J Ethnopharmacol*, 1986; 18(1): 95-101.
56. Keshri G., Singh M.M., Lakshmi V., Mehrotra B.N., Gupta D.N., Antifertility activity of *Caesalpinia decapetela*-a preliminary report. *Indian J Med Res*, 1988; 87: 377-378.
57. Mishra M., Gautam R.K., Mathur R., Evaluation of Antifertility potential of *Calotropis gigantea* LINN. in female albino rats. *J Pharm Res Opin*, 2011; 1(3): 92-93.
58. Gupta R.S., Sharma N., Dixit V.P., Calotropin A, novel compound for fertility control. *Ancient Science of life*, 1990; 9: 224-230.

59. Chinoy N.J., George S.M., Induction of functional sterility in male rats by low dose *Carica papaya* seed extracts treatment. *Acta Eur Fertil*, 1983; 14(6): 425-432.
60. Lohiya N.K., Goyal R.B., Antifertility investigation on the the crude chloroform extract of *Carica papaya* Linn. Seeds in male albino rats. *Indian J Exp Biol*, 1992; 30(11): 1051-1055.
61. Manivannan B., Mishra P.K., Pathak N., Sriram S., Bhande S.S., Panneerdos S., Ultrastructural changes in the testis and epididymis of the rats following treatment with benzene chromatographic fraction of the chloroform extract of the seeds of *Carica papaya*. *Phytother Res*, 2004; 18: 285-289.
62. Udoh P., Essien I., Udoh F., Effect of *Carica papaya* (paw paw) seeds extract on the morphology of pituitary-gonadal axis of male Wistar rats. *Phytother Res*, 2005; 19(12): 1065-1068.
63. Weniger B., Haag Berruier M., Anton R., Plants of Haiti used as antifertility agents. *J Ethnopharmacol*, 1982; 6(1): 67-84.
64. Yadav R., Jain G.C., Antifertility effect of the aqueous extracts of seeds of *Cassia fistula* in female rats. *Adv in contraception*, 1999; 15: 293-301.
65. Badwai P.P., Wangod, Bullar N., An Antispermato-genic effect of the seed extracts of *Celastrus paniculatus* in rats with reversible changes in liver. *J Ethnopharmacol*, 1990; 28(3): 293-303.
66. Dutta T., Basu U.P., Crude extract of *Centella asiatica* and products derived from its glycosides as oral antifertility agents. *Indian J Exp Biol*, 1968; 6(3): 181-182.
67. Keshri G., Lakshmi V., Singh M.M., Post-cotal contraceptive activity of some indigenous plants in rats. *Contraception*, 1998; 57: 357-360.
68. Chaturvedi M., Mali P.C., Ansari, Induction of reversible antifertility with a crude ethanol extract of *Citrullus colosynthis* Schard fruit in male rats. *Pharmacology*, 2003; 68(1): 38-48.
69. Piyachaturawat P., Glinsukon T., Chanjarunee A., Antifertility effect of *Citrus hystrix*. *J Ethnopharmacol*, 1985; 13: 105-110.
70. Vaithinathan S., Krishnamoorthy P., Ravikumar S., Investigation on the impact of lemon *Citrus limon* Seed extract on spermatogenesis. *Proc National Interactive Meet On Medicinal and Aromatic Plants (India)*, 2003; 308-310.
71. Suganthan D., Santhakumari G., Antifertility activity of indigenous preparation Ayush-47. *Indian J Med Res*, 1979; 7(3): 504-516.
72. Gupta R.S., Yadav R.K., Dixit V.P., Dobhal M.P., Antifertility studies of *Colebrookia oppositifolia* "Smith" leaf extract in male albino rats with special reference to testicular cell population dynamics. *Fitoterapia*, 2001; 236-245.
73. Almeida F.C.G., Lemonica I.P., The toxic effects of *Coleus barbatus* B. on the different periods of pregnancy in rats. *J Ethnopharmacol*, 2000; 73 (1-2): 53-60.
74. Benie T.I.A., Tahiri C., Duval J., Theiulant M.L., *Combretodendron africanum* bark extract as an antifertility agent I: Estrogenic effects in vivo and LH release cultured gonadotrope cells. *J Etnopharmacol*, 1990; 29(1): 13-23.
75. Gupta M., Pal D.K., Mazumdar U.K., Anti-steroidogenic activity of the methanolic extract of *Cuscuta rewflexa* Roxb. Stem and *Corchorus olitorius* Linn. Seed in mouse ovary. *Indian J Exp Biol*, 2003b; 41: 641-644.

76. Vijaykumar B., Sangamma I., Sharanabasappa A., Patil S.B., Antispermatic and hormonal effects of *Crotalaria juncea* L. seed extracts in male mice. *Asian J Androl*, 2004; 6(1): 67-70.
77. Gupta M., Mazumdar U.K., Vasmi M.L., Svakumar T., Kandar C.C., Anti-steroidogenic activity of the two Indian medicinal plants in mice. *J Ethnopharmacol*, 2004b; 90(1): 21-25.
78. Purohit A., Contraceptive efficacy of *Curcuma longa* (50% EtOH extract) with special emphasis on testicular population dynamics. *J Current Biosci*, 1991; 8: 129-134.
79. Mazumdar U.K., Gupta M., Pal D.K., Bhattacharya S., Induction of carbonic anhydrase by *Cuscuta reflexa* stem and *Corchorus olitorius* seed in mice. *Indian J Pharm Sci*, 2003; 65(4): 401-403.
80. EL Rahman H.A.A., Elbadry A.A., Mahmoud O.H., The effect of the aqueous extract of *Cynomorium coccineum* on the epididymal sperm pattern of rat. *Phytotherapy Res*, 1999; 13: 248-250.
81. Hu B., Sha H., Wang C., Yu D., Wu X., Proton NMR identification of Yuanhuatin an antifertility significant component. *Bopuxue Zazhi*, 1984; 5: 477-480.
82. Chu Y., Zhou M., Li Q., Bao Y., Antifertility effect of volatile oil from *Daucus carota* seeds. *Shengzhi Yu Biyun*, 1985; 5(1): 37-40.
83. Badami S., Aneesh R., Sankar S., Sathish kumar M.N., Antifertility activity of *Derris brevipes* variety *coriacea*. *J Ethnopharmacol*, 2003; 84: 99-104.
84. Prakash A.O., Potentialities of some indigenous plants of antifertility activity. *Int J Crude Drug Res*, 1986; 24(1): 19-24.
85. Woo W.S., Lee E.B., Kang S.S., Shin K.H., Antifertility principle of *Dictamnus albus* root bark. *Planta Med*, 1987; 53(5): 339-401.
86. Costa de Psquate R., Lragusa S., Circosta S., Investigation on *Dieffenbachia amoena* Gentil I: Endocrine effects and contraceptive activity. *J Ethnopharmacol*, 1984; 12(3): 293-303.
87. Bandara B.M.R., Jayasinghe L., Karunaratne V., Wannigamma G.P., Ecdysterone from stem of *Diploclisia glaucescens*. *Phytochemistry*, 1989; 28(4): 1073-1075.
88. Kithchlu S., Mehrotra P.K., Singh S., Progesterone potentiating effect of *Dipsacus mitis* D. Don for its contraceptive action in hamster. *Indian J Exp Biol*, 1999; 37(4): 402-405.
89. Keshri G., Oberoi R.M., Lakshmi V., Pandey K., Singh M.M., Contraceptive and hormonal properties of the stem bark of *Dysoxylum binectariferum* in rat and docking analysis of rohitukine, the alkaloid isolated from active chloroform soluble fraction. *Contraception*, 2007; 76(5): 400- 407.
90. Delgado N.M., Taboada R.J., Ortega H.A., Merchant L., Effects of a purified fraction from *Echeveria gibbiflora* aqueous crude extract on guinea pig spermatozoa. *Phytother Res*, 1999; 13(1): 46-49.
91. Sharma K.S., Mishra S., Mehta B.K., Antifertility activity of *Echinops echinatus* in male albino rats. *Indian J Med Sci*, 1988; 42: 23-26.
92. Seshadari C., Suganthan D., Santhakumari G., Biochemical changes in the uterus and uterine fluid of mated rats treated with Embelin-a non-steroidal oral contraceptive. *Indian J Exp Biol*, 1978; 16(11): 1187-1188.
93. Roy Chaudhary M., Chandrasekaran R., Mishra S., Embryotoxicity and teratogenicity studies of an Ayurvedic contraceptive pippaliyadi vati. *J Ethnopharmacol*, 2001; 74(2): 189-193.

94. Amonkar A.J., Trivedi G.K., Bhattacharya S.C., Structure of Kadalin, a biologically active component of Banakadali (*Ensete superbum*). *Indian J Chem*, 1978; 16B(1): 12-15.
95. Hiermann A., Bucar F., Studies of *Epilobium angustifolium* extracts on growth of accessory sexual organs in rats. *J Ethnopharmacol*, 1977; 55: 179-183.
96. Rajasekaran M., Bapna J.S., Lakshmanan S., Nair A.G.R., Antifertility effect in male rats of oleanolic acid, a triterpene from *Eugenia Jambolana*. *J Ethnopharmacol*, 1988; 24: 115-121.
97. Keshri G., Singh M.M., Lakshmi V., Kamboj V.P., Post-coital antifertility activity of *Ferula asafoetida* extract in female rats. *Pharmaceutical Biol*, 1999; 37(4): 273-276.
98. Singh M.M., Agnihorti A., Garg S.N., Agarwal S.K., Antifertility and hormonal properties of certain carotene sesquiterpenes of *Ferula jaeschkeana*. *Planta Med*, 1988; 489-494.
99. Prakash A.O., Pathak S., Mathur R., Post coital contraceptive action in rats of a hexane extract of the aerial part of *Ferula jaeschkeana*. *J Ethnopharmacol*, 1981; 34(2-3): 221-234.
100. Farooq T., Vanithakumari G., Bhuvaneshwari G., Malini T., Effects of anethole on seminal vesicle of albino rats. *Ancient Science of life*, 1991; 11: 9-11.
101. Xu R., Gao Y., Recent advances in chemical studies on the active principles from plants for fertility regulation. *Pure Appl Chem*, 1986; 58(5): 811-816.
102. Chou S.C., Ramanathan S., Matsui A., Rogers J., Isolation of saponins with antifertility activity from *Gleditsia horrida*. *Indian J Exp Biol*, 1971; 9: 503-504.
103. Elbeticha A., Organ S.A., Alkafahi A., Darmani H., Fetotoxic potentials of *Globularia Arabica* and *Globularia alypum* in rats. *J Ethnopharmacol*, 2001; 72: 215-219.
104. Choudhury P.K., Colchicine from *Gloriosa superba* L.: a substitute of *Colchicum autumnale* L. *Indian Drugs*, 1993; 33: 529-530.
105. Percy R.G., Calhoun M.C., Kim H.L., Seed Gossypol Variation within *Gossypium barbadense* L. Cotton. *Digital Library*, 1996; 36(1): 193-197.
106. Chang M.C., Gu Z., Saksena S.K., Effects of gossypol on the fertility of male rats, hamsters and rabbits. *Contraception*, 1980; 21(5): 461-469.
107. Hoshiai H., Uehara S., Nagbaiki F., Momono K., Mechanism of action of gossypol as a male contraceptive agent in rats in vivo study. *Nippon Funnin Gakkai Zasshi*, 1981; 26(4): 393-397.
108. Offiah V.N., Ezenwaka C.E., Antifertility properties of the hot aqueous extract of *Guaicagum officinale*. *Pharm Biol*, 2003; 41(6): 454-457.
109. Andhiwal C.K., Has C., Varshney R.P., Chemical and Pharmacological studies of *Heliotropium indicum*. *Indian Drugs*, 1985; 22 (11): 567-569.
110. Moundipa F.P., Kantchoung P., Kouetya N., Tant Chou J., Effects of aqueous extract of *Hibiscus macranthus* and *Basella alba* in mature rat testes function. *J Ethnopharmacol*, 1999; 65: 133-140
111. Kholkute S.D., Chatterjee S.R., Udupa K.N., Effect of *Hibiscus rosa-sinensis* Linn. On oestrus cycle and reproductive organs in rats. *Indian J Exp Biol*, 1976; 14 (6): 703-704.
112. Singh M.P., Singh R.H., Udupa K.N., Antifertility activity of benzene extract of *Hibiscus rosa-sinensis* flower on female albino rats. *Planta Med*, 1982; 44(3): 171-174.

113. Reddy C.M., Murthy D.R.K., Patil S.B., Antispermatic and androgenic activities of various extracts of *Hibiscus rosa-sinensis* in albino mice. *Indian J Exp Biol*, 1977; 35: 1170-1174.
114. Nivsarkar M., Patel M., Padh H., Bapu C., Shrivastava N., Blastocyst implantation failure in mice due to nonreceptive endometrium: endometrial alterations by *Hibiscus rosa-sinensis* leaf extract. *Contraception*, 2003; 71(3): 227-230.
115. Saluja A.J., Slantar D.D., Hormonal profile of *Hyptis-Hyptis saubealeus* Pott. *Ind J Pharm Sci*, 1983; 97-99.
116. Singh M.M., Chowdhury S.R., Kulshreshtha D.K., Antigestagenic activity of *Ixora finlaysoniana* in rat. *Contraception*, 1983; 48(2): 178-191.
117. Tafesse G., Mekonnen Y., In vivo and in-vitro Antifertility and anti-implantation properties of *Leonotis ocyimifolia* in rats. *African Journal of Traditional Complementary and alternative medicines*, 2005; 2(2): 103-112.
118. Singh M.M., Wadhwa V., Gupta D.N., Pal R., Post-cital contraceptive efficacy and hormonal profile of *Lepidium capitatum*. *Planta Med*, 1984; 50(2): 154-157.
119. Bhorla R., Kainsa S., *Leucas cephalotes* (Roth) Spreng: Review at a glance. *Int J Pharm*, 2013; 3(1): 77-81.
120. Purohit A., Vyas S.K., Antifertility effect of *Lindenbergia indica* (70% ethanol) extract. *Ancient Science of Life*, 2003; 22(3): 111-114.
121. Gaitonde B.B., Mahajan R.T., Antifertility activity of *Lygodium flexuosum*. *Indian J Med Res*, 1980; 72: 597-604.
122. Joshi B.C., Kumar S., Verma O.P., Chatterjee S.N., Antifertility effects of chronically administered *Malvaviscus concuttii* flower extract (SC) on male albino mice. *Indian J Exp Biol*, 1908; 18(6): 561-564.
123. Yuan J.L., Ding W.P., Shi J.P., Lu Z.Z., Zhou B.N., Studies on the antifertility components from *Marsdenia Koi*. *J Tongji Med Univ*, 1991; 11: 165-168.
124. Gupta M., Mazumdar U.K., Dutta I., Bhattacharya S., Mukherjee S., Studies on Antifertility activities of *Marsilea minuta* Linn. *Indian J Pharm Sci*, 2002a; 64(2): 176-178.
125. Mali P.C., Ansari A.S., Chaturvedi M., Antifertility effect of chronically administered *Martynia annua* extracts on male rats. *J Ethnopharmacol*, 2002; 82(2): 61-67.
126. Montanari T., Carvalho J.E., Dolder H., Effect of *Maynetus ilicifolia* Mart ex. Reissou spermatogenesis. *Contraception*, 1998; 57(5): 335-339.
127. Sharma N., Jacob D., Antifertility investigation and toxicological screening of the petroleum ether extract of the leaves of *Mentha arvensis* L. in the male albino mice. *J Ethnopharmacol*, 2001; 75(1): 5-12.
128. Chan W.Y., Tam P.P.L., Chot H.L., Effects of momorcharins on the mouse embryo at the early organogenesis stage. *Contraception*, 1986; 34(5): 537-544.
129. Naseem M.Z., Patil S.R., Patil S.B., Antispermatic and androgenic activities of *Momordica charantia* in albino rats. *J Ethnopharmacol*, 1998; 61: 9-16.
130. Yeung H.W., Ng T.B., Wong N.S. Li W.W., Isolation and characteristics of an abortifacient protein momorcochin from root tubers of *Momordica cochinchinensis* (family Cucurbitaceae). *Int J Pept Protein Res*, 1988; 30(1): 135-140.

131. Dhawan B.N., Dubey M.P., Mehrotra B.N., Rastogi R.P., Screening of Indian plants for biological activity part IX Indian. J Exp Biol, 1980; 18: 594.
132. Watcho P., Kamtchousing S., Sokeng S., Moundipa P.F., Reversible antispermatogenic and antifertility activities of *Mondia whitei* L. in male albino rat . Phytother Res, 2001; 15: 26-29.
133. Hahn D.W., Ericson E.W., Lai M.T., Probst A., Antifertility activity of *Montanoa tomentosa* (Zoapatle). Contraception, 1981; 23(2): 133-140.
134. Shukla S., Mathur R., Prakash A.O., Antifertility profile of the aqueous extract of *Moringa oleifera* roots. J Ethnopharmacol, 1988; 22(1): 51-62.
135. Udoh P., Ekpenyong J., Effect of *Mucuna urens* on the gonads of male guinea pigs. Phytother Res, 2001; 15: 99-102.
136. Kong Y.C., Cheng K.F., Cambie R.C., Waterman P.G., Yuehchukene; a novel indole alkaloid with anti-implantation activity. J Chem Soc Chem Commun, 1985; 2 :47-48.
137. Kong Y.C., Ng K.H., But P.P.H., Li Q., Yu S.X., Sources of anti-implantation alkaloid Yuehchukene in the genus *Murraya*. J Ethnopharmacol, 1986; 15(2): 195-200.
138. Mazumdar U.K., Gupta M., Pramanik G., Mukhopadhyay R.K., Antifertility activity of seed of *Nelumbo nucifera* in mice. Indian J Exp Biol, 1992; 30: 533-534.
139. Londinkar R.L., Srinivasreddy P., Somanathreddy P., Patil S.B., Nicotine induced inhibition of activities of accessory reproductive ducts in male rats. J Ethnopharmacol, 1988; 60: 215-221.
140. Keshri G., Singh M.M., Lakshmi V., Kamboj V.P., Post-cotal contraceptive efficacy of the seeds of *Nigella sativa* in rats. Indian J Phsiol Pharmacol, 1995; 39(1): 59-62.
141. Singh S.P., Male contraception through an indeginous plant *Ocium sanctum* Linn. Int J Androl, 1997; 20: 14-18.
142. Pant G., Panwar M.S., Rawat M.S.M., Negi D.S., Spermicidal activity and chemical examination OF *Pentapanax ieschenaultii* (DC) seem. Fruits. Herba Plant, 1988; 34(2): 161-162.
143. Rao M.V., Alice K.M., Contraceptive effects of *Phyllanthus amarus* in female mice. Phytother Res, 2001; 15: 265-267.
144. Sudhakaran S., Ramanathan B., Ganapathi A., Antifertility effects of petroleum ether extract of *Physalis minima* on female albino rats. Pharm Biol, 1999; 37(4): 269-272.
145. Shaaban A.H., Ahmed Z.F., A new spermatocidal principle from *Phytolacca Americana*. Gaz Egypt Soc Gynaecol Obstet, 1959; 9: 27-34.
146. Stolzenberg S.J., Parkhurst R.M., Spermicidal actions of extract and compounds from *Phytolacca dodecandra*. Contraception, 1975; 10(2): 135-143.
147. Sanyal S.N., *Pisum sativum* m-xylohydroquinone as an oral contraceptive. A critical review. Acta Endocrinol, 1956; 28(suppl): 72-82.
148. Sanyal S.N., Oral contraceptives; biological studies with two new compounds 2,6-dixylenomethane and 3,5-dioxylylquinol. J Med Intern Med, 1960; 139: 1-4.
149. Varshney I.P., Khanna N., Partial structure of a new saponin: Samain- D from the flower of *Pithecolobium saman* Benth. Indian J Pharm Sci, 1978; 40(2): 60-61.

150. Chowdhary A.K.A., Sushanta K.C., Khan A.K.A., Antifertility activity of *Plumbago zeylanica* Linn. Root. Indian J Med Res, 1982; 76(Suppl.): 99-101.
151. Bhagrava S.K., Effect of plumbagin on the reproductive function of male dog. Indian J Exp Biol, 1984a; 22(3): 153-156.
152. Parveen S., Dsa S., Kundra C.P., Pereira B.M., A comprehensive evaluation of the reproductive toxicity of *Quassia amra* in male rats. Reprod toxicol, 2003; 17(1):45- 50.
153. Prakash A.O., Saxena V., Shukla S., Tewari R.K., Mathur S., Gupta A., Anti-implantation activity of some indigenous plants in rats. Acta Eur Fertil, 1985; 16(6): 441-448.
154. Okwuasaba F.K., Osunkwo U.A., Ekwenchi M.M., Ekpenyonjg K., Anticonceptive and estrogenic effects of seed extract of *Ricinus communis* var. minor. J Ethnopharmacol, 1991; 34(2-3): 141-145.
155. Shivalingappa H., Satyanarayanan N.D., Purohit M.G., Antiimplantation and pregnancy interruption efficacy of *Rivea hypocratifomis* in rat. J Ethnopharmacol, 2001; 74(3): 245-249.
156. Andhiwal C.K., Has C., Varshney R.P., Antifertility screening and phytochemical investigation of *Ruellia prostrata* Poir. J Indian Chem Soc, 1986; 63 (10): 934.
157. Gebrie E., Makonnen E., Debella A., Zerihun L., Phyto-chemical screening and pharmacological evaluations for the anti-fertility effect of the methanolic root extract of *Rumex steudelii*. J Ethanopharmacol, 2005a; 96 (1-2): 139-143.
158. Gebrie E., Makonnen E., Debella A., Zerihun L., The possible mechanism for the antifertility action of methanolic root extract of *Rumex steudelii*. Afr Health Sci, 2005b; 2: 119-125.
159. Kong Y.C., Lau C.P., Wat K.H., Ng K.H., But P.P.H., Antifertility principle of *Ruta graveolens*. Planta Med, 1989; 55(2): 176-178.
160. Louw A., Swart P., De Knock S.S., Mechanism for the stabilization in vivo of the aziridine precursor 2(4pacetoxyphenyl)-2-chloro-N-methyl-ethyl-ammonium chloride by serum proteins. Biochem Pharmacol, 1997; 53: 189-197.
161. Elbeticha A., Al-Hamood M.H., Alkafahi A., Reproductive toxicity pontencial of *Salvia fruticosa* (Labiatae) in rats. J Ethnopharmacol, 1998; 61: 67-74.
162. Venma P.K., Sharma A., Mathur A., Sharma P., Effect of *Sarcostemma acidum* stem extract on spermatogenesis in male albino rats. Asian J Androl, 2002; 4(1): 43-47.
163. Jain G.K., Sarin J.P.S., Khanna N.M., Constitution of schefflerside-a spermicidal saponin from *Schefflera capitata*. Indian J Chem, 1977; 15B (12): 1139-1141.
164. Tu Z., Konno C., Waller D.P., Bingel A.S., Molyneux R.J., Identification of senecionine and senecionine- N-oxide an antifertility constituents in *Senecio vulgaris*. J Pharm Sci, 1988; 77(5): 461-463.
165. Singh S.P., Fertility control of female through *Sesbania sesban*. J Res Edu Indian Med, 1990; 9: 27-32.
166. Offiah V.N., Anyanwu I.I., Abortifacient activity of aqueous extract of *Spondias mombin* leaves. J Ethnopharmacol, 1989; 26: 317-320.
167. Melis M.S., Effects of chronic administration of *Stevia rebaudiana* on fertility in rats. J Ethnopharmacol, 1999; 67: 157-161.
168. Hiremath S.P., Rao S.H., Jain P.K., Jaya Y., Antifertility activity of *Striga lutea*-part I. Indian J physiol Pharmacol, 1990; 34(1): 23-25.

169. Hiremath S.P., Badami S., Swamy H.K.S., Patil S.B., Antiandrogenic effect of *Striga orobanchioides*. J Ethnopharmacol, 1997; 56: 55-60.
170. Hsu C.C., Dobberstein R.H., Bingel A.S., Fong H.H., Fransworth N.R., Biological and phytochemical investigation of plants XVI: *Strumpfia maritime*. J Pharm Sci, 1981; 70(6): 682-683.
171. Garg S.K., Antifertility screening of plants. Investigation on *Taxus baccata* Linn. Leaves. Indian J Med Res, 1972; 60(1): 159-163.
172. Gupta R.K., Krishnamurthy M., Parthasarathi J., Purpurin a new flavone from *Tephrosia purpurea* seeds. Phytochemistry, 1980; 19: 1264.
173. Gupta R.S., Sharma., Antifertility effect of *Tinospora cordifolia* (Willd.) stems extract in male rats. Indian J Exp Biol, 2003; 41(8): 885-889.
174. Rastogi P., Mahrotra B.N., Compendium of Indian Medicinal Plants Vols. 1-4. Central Drug Research Institute, Lucknow and Publication and Information Directorate: New Delhi, 2003.
175. Yeung H.W., Li W.W., β -Trichosanthin: a new abortifacient protein from the Chinese drug *Wangua*, *Trichosanthes cucumeroides*. Int J Pept Protein Res, 1987; 29(3): 289-292.
176. Kamal R., Yadav R., Sharma J.D., Efficacy of the steroidal fraction of fenugreek seed extract on fertility of male albino rats. Phytother Res, 1993; 7(2): 134-138.
177. Kassem A., A-Ahgbari A., Al-habori M., Al-Mamary M., Evaluation of potential antifertility effect of fenugreek seeds in male and female rabbits. Contraception, 2006; 73(3): 301-306.
178. Zhen Q.S., Ye X., Weiu Z.J., Recent progresses in research on *Tripterigium* a male antifertility plant. Contraception, 1985; 51(52): 121-129.
179. Huynh P.N., Hikim A.P., Wang K.C., Stefanovic K., Lue Y.H., Leung A., Long term effects of triptolide on spermatogenesis epididymal sperm function and fertility in male rats. J Androl, 2000; 21: 689-699.
180. Kumar D., Kumar A., Prakash O., Potential antifertility agents from plants: A comprehensive review. J Ethnopharmacol, 2012; 1: 1-32
181. Umopathy E., Antifertility effect of cowpeas on male rats. Central Afr J Med, 1991; 39(3): 1993.
182. Akbarsha M.A., Stanley A., Averal H.I., Effect of vincristine on Leydig cell and accessory reproductive organs. Current Sci, 1995; 68: 1053-1057.
183. Bhagrava S.K., Estrogenic and pregnancy interceptor effects of the flavonoids (VI-II) of *Vitex negundo* L. seeds in mice. Plant Med Phytother, 1984b; 18(2): 74-79.
184. Gerharai Vogel, Drug discovery and evaluation, Pharmacological assay. Springer 2nd ed, 2002; 1161-1183.
185. Robert A., Turner, Screening methods in pharmacology, 2009; 2: 85-194.